# External Quality Control Program for Real-Time PCR Testing in a Multi-Centre, Randomised Controlled Clinical Trial in Chagas Disease

J.C. Ramirez<sup>1</sup>, R. Parrado<sup>2</sup>, S. Villarroel<sup>2</sup>, A. de la Barra<sup>2</sup>, M. Rodriguez<sup>3</sup>, L. Irazu<sup>3</sup>, L. Garcia<sup>2</sup>, L. Ortiz<sup>4</sup>, F. Torrico<sup>5</sup>, J. Gascon<sup>6</sup>, I. Ribeiro<sup>7</sup>, A.G. Schijman<sup>1</sup>

¹Instituto de Investigaciones en Ingeniería Genética y Biología Molecular (INGEBI-CONICET), Buenos Aires, Argentina; ²Universidad Mayor de San Simón (IIBISMED-UMSS), Cochabamba, Bolivia; ³Instituto Nacional de Enfermedades Infecciosas (INEI-ANLIS), Buenos Aires, Argentina; ⁴Universidad Autónoma Juan Misael Saracho (UAJMS), Tarija, Bolivia; ⁵Fundación CEADES, Cochabamba, Bolivia; ⁶Barcelona Centre for International Health Research (CRESIB), Barcelona, Spain; ¬Drugs for Neglected Diseases initiative (DNDi), Geneva, Switzerland

#### **BACKGROUND**

- ✓ Accurate diagnostic tools as well as markers of parasitological response to treatment, are priorities in Chagas disease research and development.
- ✓ Real-Time PCR (qPCR) testing is an increasingly recommended diagnostic endpoint in clinical trials of drug candidates in Chagas disease.
- ✓ Recent studies have assessed performance characteristics of qPCR methods, but no formal external quality control program has provided performance assessment of the assays in use in clinical trials.

#### AIM

This study aimed to characterize the performance of the qPCR method as a diagnostic endpoint in the E1224 clinical trial through the analysis of an external quality control specially devised forthis purpose.

## **MATERIALS AND METHODS**

Table 1. Design of Quality Control Panels for E1224 trial.

T. cruzi	Concentration	QC Panel I	QC Panel II	QC Panel III	QC Panel IV
stocks	(par. eq./mL)	(day 0)	(3 months)	(6 months)	(9 months)
Tcla K98	0	CCP 101	CCP 208	CCP 306	CCP 401
	1	CCP 102	CCP 206	CCP 305	CCP 403
	10	CCP 103	CCP 207	CCP 307	CCP 402
	100	CCP 104	CCP 205	CCP 308	CCP 404
Tcld Sylvio X10 Cl1	0	CCP 107	CCP 201	CCP 301	CCP 407
	1	CCP 106	CCP 202	CCP 303	CCP 406
	10	CCP 105	CCP 204	CCP 302	CCP 405
	100	CCP 108	CCP 203	CCP 304	CCP 408
TcV LL014-1- R1 Cl1	0	CCP 109	CCP 213	CCP 313	CCP 409
	1	CCP 110	CCP 214	CCP 316	CCP 410
	10	CCP 112	CCP 215	CCP 315	CCP 412
	100	CCP 111	CCP 216	CCP 314	CCP 411
TcVI CL- Brener	0	CCP 116	CCP 209	CCP 311	CCP 416
	1	CCP 114	CCP 211	CCP 310	CCP 414
	10	CCP 115	CCP 210	CCP 309	CCP 415
	100	CCP 113	CCP 212	CCP 312	CCP 413

## **DNA** extraction:

300  $\mu$ L of Guanidine-EDTA-Blood samples spiked with 200 pg of an Internal Amplification Control (IAC) were processed using of High Pure PCR Template Preparation kit (Roche, USA) and eluted in 100  $\mu$ L of elution buffer.

## qPCR procedure:

Duplex qPCR targets the satellite *T. cruzi* DNA and the IAC DNA sequences. The qPCR reactions were carried out using FastStart Universal Probe Master Mix (Roche, Germany) with 5  $\mu$ L of DNA in a final volume of 20  $\mu$ L.

Cycling: 10 minutes at 95 °C 40 cycles: 15 seconds at 95

40 cycles: 15 seconds at 95 °C 1 minute at 58 °C

## qPCR devices:

ABI7500 (Applied Biosystems, USA) >>> Core Lab Rotor-Gene Q (Corbett Life Science, UK) >>> LabB

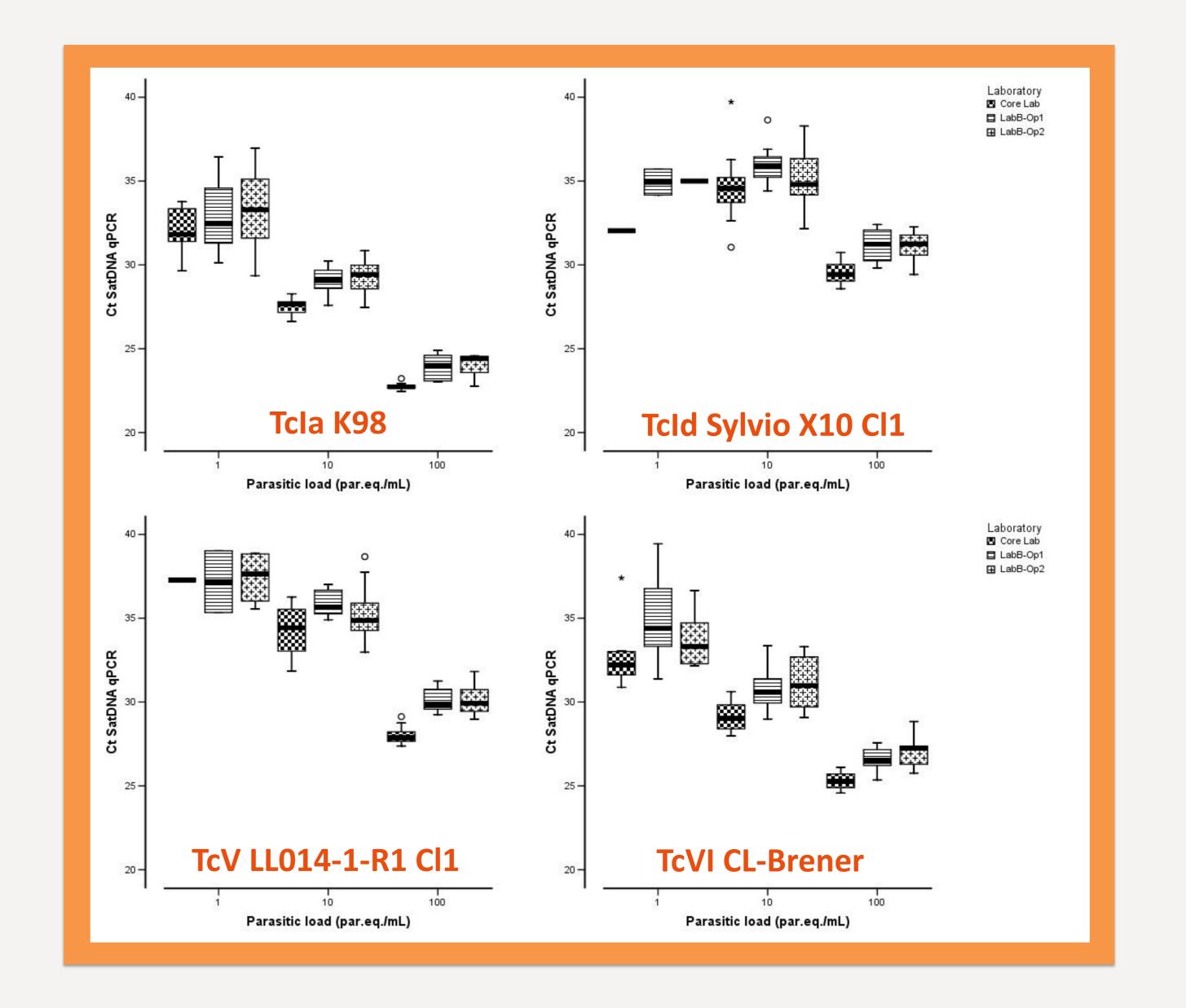
### RESULTS

Table 1. Accordance and Concordance analysis of qualitative qPCR results.

T. cruzi stocks	Laboratory	Total of	Number of Positive Replicates		
		Replicates	1 par. eq./mL	10 par. eq./mL	100 par. eq./mL
All	Core Lab	44	22	43	44
	LabB-Op1	48	26	42	48
	LabB-Op2	48	29	45	48
	Accordance	[CI95] (%)	49.8 [49.1-54.3]	86.6 [79.7-94.3]	100 [100-100]
	Concordance [CI95] (%)		50.3 [48.6-53.1]	86.7 [79.0-94.4]	100 [100-100]
	COR [CI95]		0.98 [0.96-1.18]	0.99 [0.97-1.13]	1 [1-1]
Tcla K98	Core Lab	11	11	11	11
	LabB-Op1	12	12	12	12
	LabB-Op2	12	12	12	12
	Accordance [CI95] (%)		100 [100-100]	100 [100-100]	100 [100-100]
	Concordance [CI95] (%)		100 [100-100]	100 [100-100]	100 [100-100]
	COR [CI95]		1 [1-1]	1 [1-1]	1 [1-1]
Told Sylvio X10 Cl1	Core Lab	11	1	10	11
	LabB-Op1	12	2	8	12
	LabB-Op2	12	1	11	12
	Accordance [CI95] (%)		78.1 [62.6-94.7]	71.7 [60.4-89.3]	100 [100-100]
	Concordance [CI95] (%)		79.7 [63.2-94.4]	70.3 [52.1-89.1]	100 [100-100]
	COR [	CI95]	0.91 [0.89-1.41]	1.07 [0.90-2.40]	1 [1-1]
TcV LL014- 1-R1 Cl1	Core Lab	11	1	11	11
	LabB-Op1	12	2	10	12
	LabB-Op2	12	4	10	12
	Accordance	e [CI95] (%)	66.8 [56.2-88.2]	78.6 [66.8-94.1]	100 [100-100]
	Concordance [CI95] (%)		67.2 [52.7-88.7]	79.4 [62.3-94.4]	100 [100-100]
	COR [CI95]		0.99 [0.89-2.14]	0.95 [0.91-1.41]	1 [1-1]
TcVI CL-	Core Lab	11	9	11	11
	LabB-Op1	12	10	12	12
	LabB-Op2	12	12	12	12
Brener	Accordance [CI95] (%)		79.7 [70.1-94.7]	100 [100-100]	100 [100-100]
	Concordance [CI95] (%)		78.9 [61.9-94.4]	100 [100-100]	100 [100-100]
	COR [CI95]		1.05 [0.98-1.71]	1 [1-1]	1 [1-1]

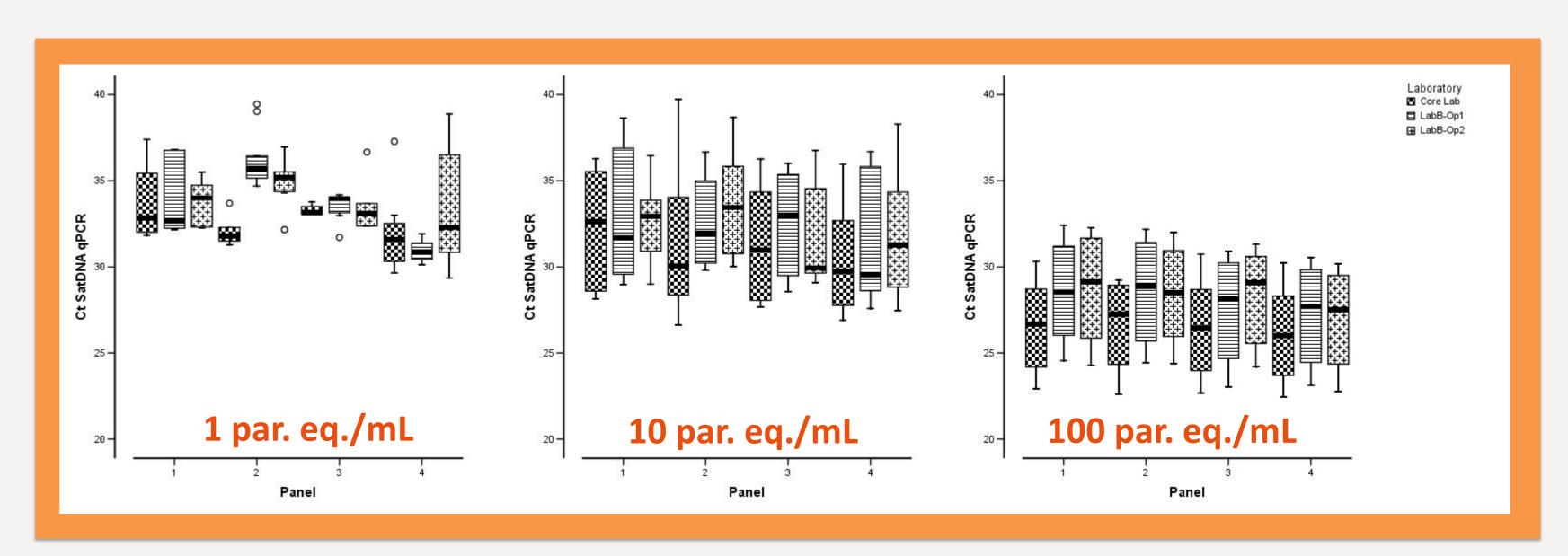
Accordance: Intra-laboratory agreement Concordance: Inter-laboratory agreement COR: Concordance Odds Ratio COR= Accordance \* (100 - Concordance) Concordance \* (100 - Accordance)

Figure 2. Analysis of QC Panels' stability after 9 months of preparation.



There were statistical differences between both laboratories but not between both operators from LabB for all *T. cruzi* stocks.

Figure 1. Analysis of inter-laboratory precision for all *T. cruzi* stocks.



## CONCLUSIONS

- ✓ There was a high within (Accordance) and between (Concordance) laboratory agreement in the qualitative results of this study, independently of the *T. cruzi* DTU and stock used.
- ✓ The results obtained for the QC panels after nine months of preparation evidenced the high stability of Guanidine-EDTA-blood samples (no statistical differences between panels).
- ✓ An External Quality Control Program for molecular diagnosis of Chagas disease is feasible and informative, allowing broader implementation of qPCR testing in clinical trial settings.













